

eCube Plasmid DNA Midi Kit

*** Things to do before starting**

1. Brief spin RNase A tube and add the RNase A to XPD1 Buffer and mix well.
Store XPD1 Buffer at 2-8°C after adding RNase A.
2. If precipitates have formed in XPD2 Buffer, warm the buffer in 37°C waterbath to dissolve precipitates.



Well-grown bacterial culture



1. Harvest the bacterial culture
Centrifugation at **6,000 x g** for **15 min**



2. Add **8ml of XPD1 Buffer (RNase A added)** to resuspend by vortexing or pipetting.
3. Add **8ml of XPD2 Buffer** and mix gently by inverting 10-15 times.
4. Add **8ml of XPD3 Buffer** and mix immediately by inverting 10-15 times.



5. Centrifuge at **15,000 x g** for **20 min** at **4°C**



* Place a XPD Midi Column into a 50 ml centrifuge tube. Add 5 ml of XEQ Buffer to equilibrate the XPD Midi column by gravity flow.



6. Transfer the supernatant to the **XPD Midi Column** and allow the column to empty by gravity flow. Discard the filtrate.
7. Add **12ml of Wash Buffer** to the XPD Midi column and allow the column to empty by gravity flow. Discard the filtrate.
8. Place the XPD Midi column into a new 50ml centrifuge tube (not provided) and add **8ml of Elution Buffer** to elute DNA by gravity flow.



9. Precipitate DNA by adding **6ml of isopropanol** to the eluted DNA.
Mix well by inverting the tube 10 times.



10. Centrifuge at **20,000 x g** for **30 min** at **4°C**



11. Carefully remove the supernatant and wash the DNA pellet with **5ml of room temperature 70% ethanol**.



12. Centrifuge at **20,000 x g** for **10 min** at **4°C**



13. Carefully remove the supernatant.
Then air-dry the DNA pellet until the tube is completely dry.
(Or incubate the DNA pellet at 70 °C for 10 min.)
14. Dissolve the DNA pellet in **300ul or a suitable volume of TE buffer or ddH₂O**.



15. You can get pure plasmid DNA.

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