

Product Insert

QuantiSpeed Probe Hi-Rox Kit

Cat. No.

QS735-02 200 x 20ul reaction: 2 x 1ml QS735-05 500 x 20ul reaction: 5 x 1ml QS735-10 1000 x 20ul reaction: 10 x 1ml

Shipping: On Dry/Blue Ice Store at -20 °C For research use only.

Storage and Stability

The QuantiSpeed Probe Hi-Rox Kit is shipped on Dry/Blue Ice. All kit components should be stored at -20°C upon receipt. Excessive freeze/thawing is not recommended. When stored under optimum conditions, the reagents are stable for a minimum of 12 months from date of purchase.

Quality Control

The QuantiSpeed Probe Hi-Rox Kit and its components are extensively tested for activity, processivity, efficiency, heat activation, and sensitivity, absence of nuclease contamination and absence of nucleic acid contamination.

Safety Precautions

Harmful if swallowed. Irritating to eyes, respiratory system and skin. Please refer to the material safety data sheet for further information.

Description

The QuantiSpeed Probe Hi-Rox Kit has been developed for fast, highly reproducible real-time PCR and has been validated on commonly used real-time instruments. The kit has been formulated for use with probe-detection technology, including TaqMan®, Scorpions® and molecular beacon

probes. A combination of the latest advances in buffer chemistry and enhancers, together with an antibodymediated hot-start DNA polymerase system, ensures that the QuantiSpeed Probe Hi-Rox Kit delivers fast, highly-specific and ultra-sensitive real-time PCR.

QuantiSpeed Probe Hi-Rox is provided as a 2x mastermix containing all the components necessary for real-time PCR, including dNTPs, stabilisers and enhancers.

Kit Components

Reagent	200x20ul	500x20ul	1000x20ul
QuantiSpeed Probe Hi-Rox Mix (2x)	2x1ml	5x1ml	10x1ml

Instrument compatibility

The QuantiSpeed Probe Hi-Rox Kit has been optimized for use with all probe chemistries, including TaqMan, FRET, Scorpions and molecular beacon probes on real-time PCR instruments listed in the following compatibility table, each of these instruments having the capacity to analyze the real-time PCR data with the passive reference signal either on or off. The kit is also compatible with several instruments that do not require the use of ROX, such as the Qiagen (Corbett) Rotor-Gene™ 6000, the Bio-Rad CFX96 or the Roche LightCycler® 480.

Manufacturer	Model	
ABI (Life Technologies)	7000, 7300, 7700, 7900, 7900HT, 7900HT FAST, StepOne™, StepOne™ Plus	

General considerations

To help prevent any carry-over DNA contamination, we recommend that separate areas are maintained for reaction setup, PCR amplification and any post-PCR gel analysis. It is essential that any tubes containing amplified PCR product are not opened in the PCR set-up area.

Primers and Probe: These guidelines refer to the design and set up of TaqMan probe-based PCR. Please refer to the relevant literature when using other probe type. The specific amplification, yield and overall efficiency of any real-time PCR can be critically affected by the sequence and concentration of the primers, as well as by the amplicon length

We strongly recommend taking the following points into consideration when designing and running your real-time PCR:

- ·use primer-design software, such as Primer3 (http://frodo.wi.mit.edu/primer3/) or visual OMPTM (http://dnasoftware.com/). Primers should have a melting temperature (Tm) of approximately 60°C; the Tm of the Probe should be approximately 10°C higher than that of the primers
- optimal amplicon length should be 80-200bp, and should not exceed 300bp
- final primer concentration of 400nM is suitable for most Probe based reactions, however to determine the optimal concentration we recommend titrating in the range 0.2-1 μ M. The forward and reverse primers concentration should be equimolar
- · a final probe concentration of 100nM is suitable for most applications; we recommend that the final probe concentration is at least two-fold lower than the primer concentration

Note: Multiplex real-time PCR probe concentrations in excess of 100nM, can result in cross channel fluorescence

Template: it is important that the DNA template is suitable for use in PCR in terms of purity and concentration. In addition, the template needs to be devoid of any contaminating PCR inhibitors (e.g. EDTA). The recommended amount of template for PCR is dependent upon the type of DNA used. The following points should be considered when using genomic DNA and cDNA templates:

- Genomic DNA: use up to 1ug of complex (e.g. eukaryotic) genomic DNA in a single PCR. We recommend using the PKT e-Cube DNA Mini Kit for high yield and purity from both prokaryotic and eukaryotic sources
- cDNA: the optimal amount of cDNA to use in a single PCR is dependent upon the copy number of the target gene. We suggest using 100ng cDNA per reaction, however it may be necessary to vary this amount. To perform a two-step RT-PCR, we recommend using the PKT cDNA Synthesis Kit for reverse transcription of the purified RNA. For high yield and purity of RNA, use the PKT e-Cube RNA Mini Kit.

MgCl₂: The QuantiSpeed Probe mix contains an optimized concentration of MgCl₂, it is not necessary to supplement the mix further.

PCR controls: It is important to detect the presence of contaminating DNA that may affect the reliability of the data. Always include a no-template control (NTC), replacing the template with PCR grade water. When performing a two-step RT-PCR, set up a no-RT control as well as an NTC for the PCR.

Optional ROX: The QuantiSpeed Probe Hi-Rox Kit is premixed with Rox (5-carboxy-rhodamine, succinymidyl ester), so that where necessary, ROX fluorescence can be optionally detected on certain real-time instruments. If your real-time instrument has the capability of using ROX and you wish to use this option, then this option must be selected by the user in the software (see notice to purchaser Trademark and Licensing Information, point4).

Procedure

Reaction mix composition:

Prepare a PCR mastermix. The volumes given below are based on a standard 20ul final reaction mix and can be scaled accordingly.

Reagent	Volume	Final concentration
2x QuantiSpeed Probe Hi-Rox Mix	10ul	1x
10uM Forward Primer	0.8ul	400nM
10uM Reverse Primer	0.8ul	400nM
10uM Probe	0.4ul	200nM
Template	<100ng cDNA, <1ug genomic	variable
H ₂ O	As required	
	20ul Final volume	

Sensitivity testing and Ct values:

When comparing QuantiSpeed with a mix from another supplier we strongly recommend amplifying from a 10-fold template dilution series. Loss of detection at low template concentration is the only direct measurement of sensitivity. An early Ct value is not an indication of good sensitivity, but rather an indication of speed.

Suggested thermal cycling conditions:

The real-time PCR conditions, in the below, are suitable for the QuantiSpeed Probe Hi-Rox Kit with the amplicons of up to 200bp. These cycling paramenters have been optimized on a number of platforms, however they can be varied to suit different machine-specific protocols.

Cycles	Temperature	Time	Notes
1	*95℃	*2-3min	Polymerase activation
	95℃	5s	Denaturation
40	60-65°C	**20-30s	Annealing/extension (acquire at end of step)

^{* 2}min for cDNA, 3min for genomic DNA

Technical Support

If the troubleshooting guide does not solve the difficulty you are experiencing, please contact Technical Support with details of reaction setup, cycling conditions and relevant date.

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^{**} Not recommended to anneal/extend beyond 30 seconds